

NeurotrypsinTechnical Field

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The present invention is directed to neurotrypsins and to a pharmaceutical composition which contains these substances or has an influence on these substances.

10 Disclosure of Invention

Neurotrypsin is a newly discovered serine protease, which is predominantly expressed in the brain and in the lungs; the expression in the brain takes place nearly exclusively in the neurons.

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Neurotrypsin has a previously not yet found domain composition: besides the protease domain, there are found 3 or 4 SRCR (scavenger receptor cysteine-rich) domains and one Kringle domain. It is to be pointed out that the combination of Kringle and SRCR domains have not yet been found in proteins. At the amino terminus of the neurotrypsin protein there is a segment of more than 60 amino acids, which has an extremely high proportion of proline and basic amino acids (arginine and histidine).

The invention is characterized by the characteristics in the independent claims. Preferred embodiments are defined in the dependent claims.

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The newly found neurotrypsins

Sub C

- neurotrypsin of the human (compound of the formula I),
- neurotrypsin of the mouse (compound of the formula II)

30 differ structurally very much from the so far known serine proteases.

The serine protease whose protease domain is structurally most closely related with the protease domain of the new compounds, namely plasmin (of the human), has only a 44 % amino acid sequence identity.

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The proline-rich, basic segment at the amino terminus has a certain resemblance with the basic segments of the netrins and the semaphorins/collapsins. Due to this

segment, it is probable that neutrotrypsin may be enriched by means of heparin-affinity chromatography.

5 The neutrotrypsins of the human (compound of the formula I) and of the mouse (compound of the formula II) exhibit a very high structural similarity among each other.

The identity of the amino acid sequences of the native proteins of the compounds of the formulas I or II amounts to 81%.

Sub C2 The neutrotrypsin of the human (compound of the formula I) has a coding sequence of 2625 nucleotides. The coded peptide of the compound of the formula I has a length of 875 amino acids and contains a signal peptide of 20 amino acids. The neutrotrypsin of the mouse (compound of the formula II) has a coding sequence of 2283 nucleotides. The coded protein of the compound of the formula II has a length of 761 amino acids and contains a signal peptide of 21 amino acids. The reason for the greater length of the neutrotrypsin of the human consists therein that the human neutrotrypsin has 4 SRCR domains, whereas the neutrotrypsin of the mouse has only 3 SRCR domains.

20 The domains which are present in both compounds (compound of the formula I and compound of the formula II) have a high degree of sequence similarity. The corresponding SRCR domains of the compounds of the formulas I and II have an amino acid sequence identity from 81% to 91%. The corresponding Kringle domains have an amino acid sequence identity of 75%. A high degree of similarity consists also in the enzymatically active (i.e. proteolytic) domain (90% amino acid sequence identity).

25 The protease domains of the neutrotrypsins of the human (compound of the formula I) and of the mouse (compound of the formula II) are aligned in the following section, in order to illustrate the high degree of sequence identity.

CGLRLLHRRQKRIIGGKNSLRGGWPWQVSLRLKSSHGDGRLLCGATLLSS	50
. : . : .	
CGLRLLHRRQKRIIGGNNSLRGAWPWQASLRLRSAHGDGRLLCGATLLSS	
CWVLTAAHCFKRYGNSTRSYAVRVG DYHTLVPEEFEEEIGVQQIVI HREY	100
.	
CWVLTAAHCFKRYGNNSRSYAVRVG DYHTLVPEEFEQEIGVQQIVI HRNY	
RPDRSDYDIALVRLQGP EEQCARFSSHVLPACLPLWRERPQKTASN CYIT	150
: : .	
RPDRSDYDIALVRLQGPGEQCARLSTHVLPACLPLWRERPQKTASN CHIT	
GWGDTGRAY SRTLQQAAIPLLPKRFCEERYKGRFTGRMLCAGNLHEHKRV	200
: .	
GWGDTGRAY SRTLQQAAVPLL PKRFCKERYKGLFTGRMLCAGNLQEDNRV	
DSCQGDGGPLMCERPGESVVYGVTSWGYGCGVKDSPGVYTKVSAFVPW	250
:	
DSCQGDGGPLMCEKPDESVVYGVTSWGYGCGVKDTPGVYTRVPAFVPW	
I KSVTKL	258
.	
I KSVTSL	

Sub C3 From the 258 amino acid sequence positions included in the comparison there are 233 amino acids that are identical in both compounds (upper sequence: compound of the formula I; lower sequence: compound of the formula II; identical amino acids are indicated by vertical lines).

The inventive neurotrypsins are unique when compared with the known serine proteases in that they are expressed according to currently available observations in a distinct degree in neurons. A further organ with a strong expression of neurotrypsin are 10 the lungs (see Gschwend et al., Mol. Cell. Neurosci. 9, pages 207-219, 1997).

The proteins that are structurally most similar to the compounds of the formulas I or II are serine proteases, such as tissue-type plasminogen activator (tPA), urokinase-type plasminogen activator (uPA), plasmin, trypsin, apolipoprotein (a), coagulation factor XI, neuropsin, and acrosin.

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In the adult brain, the inventive compounds are expressed predominantly in the cerebral cortex, the hippocampus, and the amygdala.

10 In the adult brain stem and the spinal cord, the inventive compounds are expressed predominantly in the motor neurons. A slightly weaker expression is found in the neurons of the superficial layers of the dorsal horn of the spinal cord.

15 In the adult peripheral nervous system, the inventive compounds are expressed in a subpopulation of the sensory ganglia neurons.

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The inventive compounds were found in connection with a study aimed at discovering trypsin-like serine proteases in the nervous system.

20 The first compound that was found and characterized was the compound of the formula II (Gschwend et al., Mol. Cell. Neurosci. 9, pages 207-219, 1997).

25 By means of an alignment of the protease domains of 7 known serine proteases (tissue-type plasminogen activator, urokinase-type plasminogen activator, thrombin, plasmin, trypsin, chymotrypsin, and pancreatic elastase) in the proximity of the histidine and the serine of the catalytic triade of the active site, the sequences of the so-called primer oligonucleotides for the polymerase chain reaction were determined.

30 The primer oligonucleotides were used in a polymerase chain reaction (PCR) together with ss-cDNA from total RNA of the brains of 10 days old mice and resulted in the amplification of a cDNA fragment of a length of approximately 500 base pairs.

35 This cDNA fragment was used successfully for the isolation of further cDNA fragments by screening commercially available cDNA libraries. Together, the isolated cDNA fragments covered the full length of the coding part of the compound of the formula II.

By conventional DNA sequencing the complete nucleotide sequence and the amino acid sequence deduced therefrom was obtained.

5 The compound of the formula I was cloned based on its pronounced similarity with the compound of the formula II.

The primer oligonucleotides used were synthesized according to the known sequence of the compound of the formula II.

10 The cloning of the compound of the formula I was performed by means of two commercially available cDNA libraries from fetal human brain.

This procedure for the cloning can also be used for the isolation of the homologous compounds of other species, such as rat, rabbit, guinea pig, cow, sheep, pig, primates, birds, zebra fish (*Brachydanio rerio*), *Drosophila melanogaster*, *Caenorhabditis elegans* etc.

20 The coding nucleotide sequences can be used for the production of proteins with the coded amino acid sequences of the compounds of the formulas I or II. A procedure developed in our laboratory allows the production of recombinant proteins in myeloma cells as fusion proteins with an immunoglobulin domain (constant domain of the kappa light chain). The principle of the construction is given in detail by Rader et al. (Rader et al., *Eur. J. Biochem.* 215, pages 133-141, 1993). The fusion protein produced by the 25 myeloma cells was isolated by immunoaffinity chromatography using a monoclonal antibody against the Ig domain of the kappa light chain. With the same expression method, also the native protein of a compound, starting from the coding sequence, can be produced.

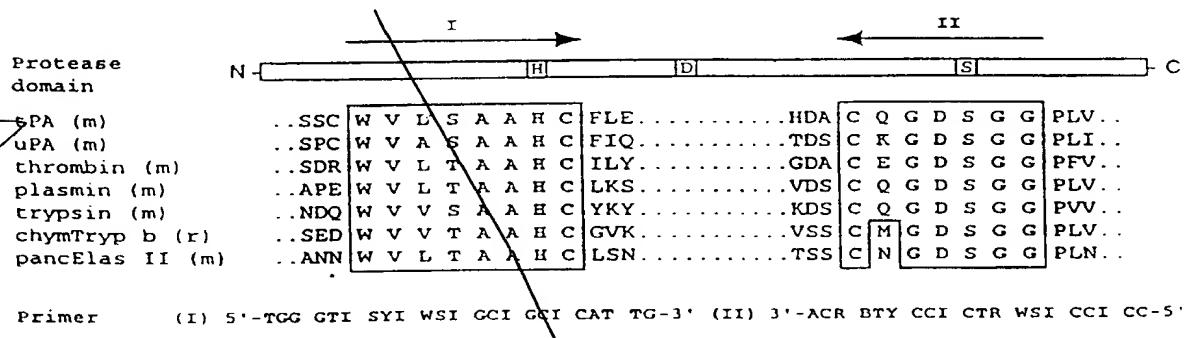
30 The coding sequences of the compounds of the formulas I or II can be used as starting compounds for the discovery and the isolation of alleles of the compounds of the formulas I or II. Both the polymerase chain reaction and the nucleic acid hybridization can be used for this purpose.

The coding sequences of the compounds of the formulas I or II can be used as starting compounds for so-called "site-directed mutagenesis", in order to generate nucleotide sequences coding the coded proteins that are defined by the compounds of the formulas I or II, or parts thereof, but whose nucleotide sequence is degenerated with respect to the compounds of the formulas I or II due to use of alternative codons.

The coding sequences of the compounds of the formulas I or II can be used as starting compounds for the production of sequence variants by means of so-called site-directed mutagenesis.

Best Modes for Carrying out the Invention (Examples)cDNA cloning of the compound of the formula II (neurotrypsin of the mouse)

5 Total RNA was isolated from the brains of 10 days old mice (ICR-ZUR) according to the method of Chomczynski and Sacchi (1987). The production of single stranded cDNA was carried out using oligo(dT) primer and a RNA-dependent DNA polymerase (SuperScript RNase H-Reverse Transcriptase; Gibco BRL, Gaithersburg, MD) according to the instruction of the supplier. For the realization of the polymerase chain reaction one forward primer was synthesized based on the amino acid sequence of the region of the conserved histidine of the catalytic triade and one primer in the backward direction was synthesized based on the amino acid sequence of the region of the conserved serine of the catalytic triade of the serine proteases. The amino acid sequences used for the determination of the oligonucleotide primers were taken from seven known serine proteases. They are presented in the following.



20 The protease domains of 7 known serine proteases (tissue-type plasminogen activator, urokinase-type plasminogen activator, thrombin, plasmin, trypsin, chymotrypsin, and pancreatic elastase) were aligned in the region of the conserved histidine and serine of the catalytic triade of the active site. The conserved amino acids of these regions were taken as the basis for the determination of the degenerated primers. The primer sequences are given according to the recommendation of the IUB nomenclature (Nomenclature Committee 1985).

25 The primers used in the PCR contained restriction sites for *Eco*RI and *Bam*HI at their 5' ends in order to facilitate a subsequent cloning.

The following primers were used:

In the reading direction (sense primers):

Sense CS
5'-GGGGAAATTCTGGGT(C/G)(T/C)(T/A)(G/C)IGCIGCICA(T/C)TG-3'

In the counter direction (antisense primers):

5'-GGGGATCCCCICCI(G/C)(A/T)(A/G)TCICC(C/T)T(G/C/T)(G/A)CA-3'.

The polymerase chain reaction was carried out under standard conditions using the DNA polymerase AmpliTaq (Perkin Elmer) according to the recommendations of the producer. The following PCR profile was employed: 93°C for 3 minutes, followed by 35 cycles of 93°C for 1 minute, 48°C for 2 minutes, and 72°C for 2 minutes. Following the last cycle, the incubation was continued at 72°C for further 10 minutes.

The amplified fragments had an approximate length of 500 base pairs. They were cut with *Eco*RI and *Bam*HI and inserted in a Blue Script vector (Bluescript SK(-), Stratagene). The resulting clones were analyzed by DNA sequence determination using the dideoxy chain termination method (Sanger et al., Proc. Natl. Acad. Sci. USA 77, pages 2163-2167, 1977) on an automated DNA sequencer (LI-COR, model 4000L; Lincoln, NE) using a commercial sequencing kit (SequiTerm long-read cycle sequencing kit-LC; Epicentre Technologies, Madison, WI). The analysis yielded a sequence of 474 base pairs of the catalytic region of the serine protease domain of the compound of the formula II.

The 474 base pair long PCR fragment was used for screening of an oligo(dT)-primed Uni-ZAP-XR cDNA library from the brain of 20 days old mice (Stratagene; cat. no. 937 319). A total of 3×10^6 lambda plaques were screened under high stringent conditions (Sambrook et al., Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory Press, 1989) using a radioactively labeled PCR fragment as a probe and 24 positive clones were found.

From the positive Lambda-Uni-ZAP-XR phagemid clones the corresponding Bluescript plasmid was cut out by *in vivo* excision according to a standard method recommended by the producer (Stratagene). In order to determine the length of the inserted fragments the corresponding Bluescript plasmid clones were digested with *Sac*I and *Kpn*I. The clones containing the longest fragments were analyzed by DNA

sequencing (as described above) and for subsequent data analysis the GCG software (version 8.1, Unix; Silicon Graphics, Inc.) was used.

Because none of the clones contained the coding sequence in full length, a second
5 cDNA library was screened. The library used in this screen was an oligo(dT)- and
random-primed cDNA library in a Lambda phage (Lambda gt10) which was based on
mRNA from 15 days old mouse embryos (oligo(dT)- and random-primed Lambda gt10
cDNA library; Clontech, Palo Alto, CA; cat. no. ML 3002a). As a probe a radioactively
labeled DNA fragment (Aval/AatII) from the 5' end of the longest clone of the first screen
10 was used and approximately 2×10^6 plaques were screened. This screen resulted in 14
positive clones. The cDNA fragments were excised with EcoRI and cloned into the
Bluecript vector (KS(+); Stratagene). The sequence analysis was carried out as
described above.

15 In this way the nucleotide sequence over the full length cDNA of 2361 and 2376
base pairs, respectively, of the compound of the formula II was obtained. With the
described procedure of PCR cloning it is possible to find and isolate also variant forms of
the compounds of the formulas I or II, as for example their alleles or their splice variants.
The described method of screening of a cDNA library allows also the detection and the
20 isolation of compounds which hybridize under stringent conditions with the coding
sequences of the compounds of the formulas I or II.

Cloning of the cDNA of the compound of the formula I (neurotrypsin of the human)

The cloning of the cDNA of the compound of the formula I was carried out basing
5 on the nucleotide sequence of the compound of the formula II. As a first step, a fragment
of the compound of the formula I was amplified using the polymerase chain reaction
(PCR). As a matrix we used the DNA obtained from a cDNA library from the brain of a
human fetus (17th - 18th week of pregnancy) which is commercially available (Oligo(dT)-
and random-primed, human fetal brain cDNA library in the Lambda ZAP II vector, cat.
10 no. 936206, Stratagene). The synthetic PCR primers contained restriction sites for
*Hind*III and *Xhol* at the 5' end in order to facilitate the subsequent cloning.

In the reading direction (sense primers):

SUN C 65
5'-GGGAAGCTTGGICA(A/G)TGGGGIACI(A/G)TITG(C/T)GA(C/T)-3'

In the counter direction (antisense primers):

5'-GGGCTCGAGCCCCAACCTGTTATGTAAIAGTTG-3'

The PCR was carried out under standard conditions using the DNA polymerase
20 AmpliTaq (Perkin Elmer) according to the recommendations of the producer. The resulting fragment of 1116 base pairs was inserted into the Bluescript vector (Bluescript SK(-), Stratagene). A 600 base pairs long *Hind*III/*Stu*I fragment, corresponding to the 5'
half the 1116 base pairs long PCR fragment, was used for the screening of a Lambda
cDNA library from human fetal brain (Human Fetal Brain 5'-STRETCH PLUS cDNA
25 library; Lambda gt10; cat. no. HL 3003 a; Clontech). 2x10⁶ Lambda plaques were
screened under high stringent conditions (Sambrook et al., Molecular Cloning: A
laboratory manual, Cold Spring Harbor Laboratory Press, 1989) by means of a
radioactively labeled PCR fragment, and 23 positive clones were found and isolated.

30 From the positive Lambda gt10 clones the corresponding cDNA fragments were
excised with *Eco*RI and inserted into a Bluescript vector (Bluescript KS(+), Stratagene).
The sequencing was carried out by means of the dideoxy chain termination method
(Sanger et al., Proc. Natl. Acad. Sci. USA 77, pages 2163-2167, 1977), using a
commercial sequencing kit (SequiTherm long-read cycle sequencing kit-LC; Epicentre
35 Technologies, Madison, WI) and Bluescript-specific primers.

In an alternative sequencing strategy, the cDNA fragments of the positive Lambda gt10 clones were PCR amplified using Lambda-specific primers. The sequencing was carried out as described above.

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The computerized analysis of the sequences was performed by means of the program package GCG (version 8.1, Unix; Silicon Graphics Inc.).

In this way the nucleotide sequence over the full length of the cDNA of 3350 base pairs was obtained. With the described procedure for PCR cloning it is possible to find and to isolate also variant forms of the compounds of the formulas I or II, as for example their alleles or their splice variants. The described procedure for the screening of a cDNA library allows also the discovery and the isolation of compounds which hybridize under stringent conditions with the coding sequences of the compounds of the formulas I
10 or II.
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Visualization of the coded sequences of the compounds of the formulas I or II by means of antibodies

line C1 The more than 60 amino acids long proline-rich, basic segment at the amino terminus of the coded sequence of the compounds of the formulas I or II is well suited for the production of antibodies by means of synthesizing peptides and using them for immunization. We have selected two peptide sequences with a length of 19 and 13 amino acids from the proline-rich, basic segment at the amino terminus of the coded sequence of the compound of the formula II for the generation of antibodies. The peptides had the following sequences:

Peptide 1: H₂N-SRS PLH RPH PSP PRS QX-CONH₂

Peptide 2: H₂N-LPS SRR PPR TPR F-COOH

The two peptides were synthesized chemically, coupled to a macromolecular carrier (Keyhole Limpet Hemocyanin), and injected into 2 rabbits for immunization. The resulting antisera exhibit a high antibody titer and could successfully be used both for the identification of native neurotrypsin in brain extract of the mouse and for the identification of recombinant neurotrypsin. The employed procedure for the generation of antibodies can also be used for the generation of antibodies against the coded sequence of the compound of the formula I.

The resulting antibodies against the partial sequences of the coded sequences of the compounds of the formulas I or II can be used for the detection and the isolation of variant forms of the compounds of the formulas I or II, as for example alleles or splice variants. Such antibodies can also be used for the detection and isolation of gene technologically generated variants of the compounds of the formulas I or II.

Purification of the coded sequences of the compounds of the formulas I or II

Besides conventional chromatographic methods, as for example ion exchange chromatography, the purification of the coded sequences of the compounds of the formulas I or II can also be achieved using two affinity chromatographic purification procedures. One affinity chromatographic purification procedure is based on the availability of antibodies. By coupling the antibodies on a chromatographic matrix, a purification procedure results, in which a very high degree of purity of the corresponding compound can be achieved in one step.

Another important feature that can be used for the purification of the coded sequences of the compounds of the formulas I or II is the proline-rich, basic segment at the amino terminus. It may be expected that, due to the high density of positive charges, this segment mediates the binding of the coded sequences of the compounds of the formulas I or II to heparin and heparin-like affinity matrices. This principle allows also the isolation, or at least the enrichment, of variant forms of the coded sequences of the compounds of the formulas I or II, as for example their alleles or splice variants. Likewise the heparin affinity chromatography can be used for the isolation, or at least the enrichment, of species-homologous proteins of the compounds of the formulas I or II.

Industrial Applicability

The coding sequences of the formulas I and II can be used for the production of the coded proteins or parts thereof of the formulas I and II. The production of the coded proteins can be achieved in procaryotic or eucaryotic expression systems.

The gene expression pattern of the inventive compounds in the brain is extremely interesting, because these molecules are expressed in the adult nervous system predominantly in neurons of those regions that are thought to play an important role in learning and memory functions. Together with the recently found evidence for a role of extracellular proteases in neural plasticity, the expression pattern allows the assumption that the proteolytic activity of neutrotrypsin has a role in structural reorganizations in connection with learning and memory operations, for example operations which are involved in the processing and storage of learned behaviors, learned emotions, or memory contents. The inventive compounds may, thus, represent a target for pharmaceutical intervention in malfunctions of the brain.

The gene expression pattern of the inventive compounds in the cerebral cortex (especially layers V and VI) is extremely interesting, because a reduction of the cellular differentiation in the cerebral cortex has been found to be associated with schizophrenia. The inventive compounds may, thus, be a target for pharmaceutical intervention in schizophrenia and related psychiatric diseases.

The coding sequences of the inventive compounds have been found to be increased in the neurons located adjacent to the damaged tissue of a focal ischemic stroke, indicating that the inventive compounds play a role in the tissue reaction in the injured cerebral tissue. The inventive compounds may, thus, represent a target for pharmaceutical intervention after ischemic stroke and other forms of neural tissue damage.

Tissue-type plasminogen activator, a serine protease related to the inventive compounds, has recently been found to be involved in excitotoxicity-mediated neuronal cell death. A similar function is conceivable for the inventive compounds and, thus, the inventive compounds represent a possible target for a pharmacological intervention in diseases in which cell death occurs.

The gene expression pattern of the inventive compounds in the spinal cord and in the sensory ganglia is interesting, because these molecules are expressed in the adult nervous system in neurons of those brain regions that are thought to play a role in the processing of pain, as well as in the pathogenesis of pathological pain. The inventive compounds may, thus, be a target for pharmaceutical intervention in pathological pain.

10 In the following part statements concerning the compounds of the formulas I or II
are given:

- 16 -

(1) INFORMATION ABOUT THE COMPOUND OF THE FORMULA I

(Neurotrypsin of the human)

(i) SEQUENCE CHARACTERISTICS:

5 (A) LENGTH: 3350 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single strand
(D) TOPOLOGY: linear

10 (ii) MOLECULE TYPE: cDNA to mRNA

15 (vi) ORIGINAL SOURCE:

(A) ORGANISM: Homo sapiens
(D) DEVELOPMENT STAGE: fetal
(F) TISSUE TYPE: brain

20 (vii) IMMEDIATE SOURCE:

(A) LIBRARY: human fetal brain 5'-stretch plus cDNA library in the lambda gt10 vector; catalog No. HL 3003a; Clontech, Palo Alto, CA, USA.
25 (B) CLONE: cDNA Clone No.:
3-1, 3-2, 3-6, 3-7, 3-8, 3-10, 3-11, 3-12

30 (ix) FEATURE:

(A) NAME/KEY: Signal peptide
(B) LOCATION: 44 .. 103

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5 (ix) FEATURE:

(A) NAME/KEY: mature peptide
(B) LOCATION: 104 .. 2668

10

(ix) FEATURE:

(A) NAME/KEY: coding sequence
(B) LOCATION: 44 .. 2668

15 (ix) FEATURE:

(A) NAME/KEY: Proline-rich, basic segment
(B) LOCATION: 104 .. 319

20 (ix) FEATURE:

(A) NAME/KEY: Kringle domain
(B) LOCATION: 320 .. 538

25 (ix) FEATURE:

(A) NAME/KEY: SRCR domain 1
(B) LOCATION: 551 .. 856

30

(ix) FEATURE:

(A) NAME/KEY: SRCR domain 2
(B) LOCATION: 881 .. 1186

35

(ix) FEATURE:

(A) NAME/KEY: SRCR domain 3

5 (B) LOCATION: 1202 .. 1504

(ix) FEATURE:

10 (A) NAME/KEY: SRCR domain 4

(B) LOCATION: 1541 .. 1846

15 (ix) FEATURE:

(A) NAME/KEY: proteolytic domain

(B) LOCATION: 1898 .. 2668

20 (ix) FEATURE:

(A) NAME/KEY: histidine of the catalytic triade

(B) LOCATION: 2069 - 2071

25

(ix) FEATURE:

(A) NAME/KEY: aspartic acid of the catalytic triade

(B) LOCATION: 2219 - 2221

30

(ix) FEATURE:

(A) NAME/KEY: serine of the catalytic triade

35 (B) LOCATION: 2516 .. 2518

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(ix) FEATURE:

5 (A) NAME/KEY: polyA signal
 (B) LOCATION: 2873 .. 2878

(ix) FEATURE

10 (A) NAME/KEY: polyA signal
 (B) LOCATION: 3034 .. 3039

15 (ix) FEATURE:

 (A) NAME/KEY: polyA signal
 (B) LOCATION: 3215 .. 3220

20 (ix) FEATURE:

 (A) NAME/KEY: 3'UTR
 (B) LOCATION: 2669 .. 3350

25

(ix) FEATURE

30 (A) NAME/KEY: 5'UTR
 (B) LOCATION: 1 .. 43

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Compound of the formula I (neurotrypsin of the human)

CGGAAGCTGG	GGAGCATGGA	CCAGACCCCCG	CAGCGCTGGC	ACC	ATG	ACG	CTC	GCC	55
					Met	Thr	Leu	Ala	
					-20				
CGC	TTC	GTG	CTA	GCC	CTG	ATG	TTA	GGG	103
Arg	Phe	Val	Leu	Ala	Leu	Met	Leu	Gly	
-15						-10		-5	-1
TTT	GAT	TCT	GTC	CTC	AAT	GAT	TCC	CTC	151
Phe	Asp	Ser	Val	Leu	Asn	Asp	Ser	Leu	
1						5		10	15
CCC	CCT	GCG	GGT	CCG	CAC	TAC	CCC	TAT	199
Pro	Pro	Ala	Gly	Pro	His	Tyr	Pro	Tyr	
20						25			30
CCC	CCG	ACG	ACG	CGT	CCG	CCG	CCT	CTC	247
Pro	Pro	Thr	Thr	Arg	Pro	Pro	Pro	Pro	
35						40			45
CCG	CGG	GCG	CTC	CCT	GCC	CAG	CCC	ACC	295
Pro	Arg	Ala	Leu	Pro	Ala	Gln	Arg	Pro	
50						55		60	
ACG	CCC	CGG	CCG	CAC	CCC	TGG	GGC	TGC	343
Thr	Pro	Arg	Pro	His	Pro	Trp	Gly	Cys	
65						70		75	80
AGC	GTG	ACG	GAC	TTC	GGC	CCC	TGT	CTG	391
Ser	Val	Thr	Asp	Phe	Gly	Ala	Pro	Cys	
85						90			95
CCC	TTC	CTG	GAG	CGG	TEG	CCC	CCA	GAG	439
Pro	Phe	Leu	Glu	Arg	Ser	Pro	Pro	Ala	
100						105		110	
CAG	CGC	CAC	AAC	TTT	TGT	CGG	TGG	GCG	487
Gln	Arg	His	Asn	Phe	Cys	Arg	Ser	Pro	
115						120		125	
TGT	TTC	TAC	GGA	GAC	GCC	CGT	GGC	AAG	535
Cys	Phe	Tyr	Gly	Asp	Ala	Arg	Gly	Lys	
130						135		140	
TGC	AGA	CAC	GGA	TCA	GTA	CGA	CTT	CGT	583
Cys	Arg	His	Gly	Ser	Val	Arg	Leu	Arg	
145						150		155	160
GGC	ACA	GTG	GAA	GTA	TAT	GCA	AGT	GGG	631
Gly	Thr	Val	Glu	Val					
165						170			175
AGC	CAC	TGG	GAT	GAT	TCT	GAT	GCA	ATT	679
Ser	His	Trp	Asp	Asp	Ser	Asp	Ala	Val	
180						185			190

CTG GGA GGA AAA GGA ATA GCA AAA CAA ACC CCG TTT TCT GGA CTG GGC Leu Gly Gly Lys Gly Ile Ala Lys Gln Thr Pro Phe Ser Gly Leu Gly 195 200 205	727
CTT ATT CCC ATT TAT TGG AGC AAT GTC CGT TGC CGA GGA GAT GAA GAA Leu Ile Pro Ile Tyr Trp Ser Asn Val Arg Cys Arg Gly Asp Glu Glu 210 215 220	775
AAT ATA CTG CTT TGT GAA AAA GAC ATC TGG CAG GGT GGG GTG TGT CCT Asn Ile Leu Leu Cys Glu Lys Asp Ile Trp Gln Gly Gly Val Cys Pro 225 230 235 240	823
CAG AAG ATG GCA GCT GCT GTC ACG TGT AGC TTT TCC CAT GGC CCA ACG Gln Lys Met Ala Ala Val Thr Cys Ser Phe Ser His Gly Pro Thr 245 250 255	871
TTC CCC ATC ATT CGC CTT GCT GGA GGC AGC AGT GTG CAT GAA GGC CGG Phe Pro Ile Ile Arg Leu Ala Gly Gly Ser Ser Val His Glu Gly Arg 260 265 270	919
GTG GAG CTC TAC CAT GCT GGC CAG TGG GGA ACC GTT TGT GAT GAC CAA Val Glu Leu Tyr His Ala Gly Gln Trp Gly Thr Val Cys Asp Asp Gln 275 280 285	967
TGG GAT GAT GCC GAT GCA GAA GTG ATC TGC AGG CAG CTG GGC CTC AGT Trp Asp Asp Ala Asp Ala Glu Val Ile Cys Arg Gln Leu Gly Leu Ser 290 295 300	1015
GGC ATT GCC AAA GCA TGG CAT CAG GCA TAT TTT GGG GAA GGG TCT GGC Gly Ile Ala Lys Ala Trp His Gln Ala Tyr Phe Gly Glu Gly Ser Gly 305 310 315 320	1063
CCA GTT ATG TTG GAT GAA GTA CGC TGC ACT GGG AAT GAG CTT TCA ATT Pro Val Met Leu Asp Glu Val Arg Cys Thr Gly Asn Glu Leu Ser Ile 325 330 335	1111
GAG CAG TGT CCA AAG AGC TCC TGG GGA GAG CAT AAC TGT GGC CAT AAA Glu Gln Cys Pro Lys Ser Ser Trp Gly Glu His Asn Cys Gly His Lys 340 345 350	1159
GAA GAT GCT GGA GTG TCC TGT ACC CCT CTA ACA GAT GGG GTC ATC AGA Glu Asp Ala Gly Val Ser Cys Thr Pro Leu Thr Asp Gly Val Ile Arg 355 360 365	1207
CTT GCA GGT GGG AAA GGC AGC CAT GAG GGT CGC TTG GAG GTA TAT TAC Leu Ala Gly Gly Lys Ser His Glu Gly Arg Leu Glu Val Tyr Tyr 370 375 380	1255
AGA GGC CAG TGG GGA ACT GTC TGT GAT GAT GGC TGG ACT GAG CTG AAT Arg Gly Gln Trp Gly Thr Val Cys Asp Asp Gly Trp Thr Glu Leu Asn 385 390 395 400	1303
ACA TAC GTG GTT TGT CGA CAG TTG GGA TTT AAA TAT GGT AAA CAA GCA Thr Tyr Val Val Cys Arg Gln Leu Gly Phe Lys Tyr Gly Lys Gln Ala 405 410 415	1351
TCT GCC AAC CAT TTT GAA GAA AGC ACA GGG CCC ATA TGG TTG GAT GAC Ser Ala Asn His Phe Glu Glu Ser Thr Gly Pro Ile Trp Leu Asp Asp 420 425 430	1399

GTC AGC TGC TCA GGA AAG ACC AGA TTT CTT CAG TGT TCC AGG CGA Val Ser Cys Ser Gly Lys Glu Thr Arg Phe Leu Gln Cys Ser Arg Arg 435 440 445	1447
CAG TGG GGA AGG CAT GAC TGC AGC CAC CGC GAA GAT GTT AGC ATT GCC Gln Trp Gly Arg His Asp Cys Ser His Arg Glu Asp Val Ser Ile Ala 450 455 460	1495
TGC TAC CCT GGC GGC GAG GGA CAC AGG CTC TCT CTG GGT TTT CCT GTC Cys Tyr Pro Gly Gly His Arg Leu Ser Leu Gly Phe Pro Val 465 470 475 480	1543
AGA CTG ATG GAT GGA GAA AAT AAG AAA GAA GGA CGA GTG GAG GTT TTT Arg Leu Met Asp Gly Glu Asn Lys Lys Glu Gly Arg Val Glu Val Phe 485 490 495	1591
ATC AAT GGC CAG TGG GGA ACA ATC TGT GAT GAT GGA TGG ACT GAT AAG Ile Asn Gly Gln Trp Gly Thr Ile Cys Asp Asp Gly Trp Thr Asp Lys 500 505 510	1639
GAT GCA GCT GTG ATC TGT CGT CAG CTT GGC TAC AAG GGT CCT GCC AGA Asp Ala Ala Val Ile Cys Arg Gln Leu Gly Tyr Lys Gly Pro Ala Arg 515 520 525	1687
GCA AGA ACC ATG GCT TAC TTT GGA GAA GGA AAA GGA CCC ATC CAT GTG Ala Arg Thr Met Ala Tyr Phe Gly Glu Gly Lys Gly Pro Ile His Val 530 535 540	1735
GAT AAT GTG AAG TGC ACA GGA AAT GAG AGG TCC TTG GCT GAC TGT ATC Asp Asn Val Lys Cys Thr Gly Asn Glu Arg Ser Leu Ala Asp Cys Ile 545 550 555 560	1783
AAG CAA GAT ATT GGA AGA CAC AAC TGC CGC CAC AGT GAA GAT GCA GGA Lys Gln Asp Ile Gly Arg His Asn Cys Arg His Ser Glu Asp Ala Gly 565 570 575	1831
GTG ATT TGT GAT TAT TTT GGC AAG AAG GCC TCA GGT AAC AGT AAT AAA Val Ile Cys Asp Tyr Phe Gly Lys Lys Ala Ser Gly Asn Ser Asn Lys 580 585 590	1879
GAG TCC CTC TCA TCT GTT TGT GGC TTG AGA TTA CTG CAC CGT CGG CAG Glu Ser Leu Ser Ser Val Cys Gly Leu Arg Leu Leu His Arg Arg Gln 595 600 605	1927
AAG CGG ATC ATT GGT GGG AAA AAT TCT TTA AGG GGT GGT TGG CCT TGG Lys Arg Ile Ile Gly Gly Lys Asn Ser Leu Arg Gly Gly Trp Pro Trp 610 615 620	1975
CAG GTT TCC CTC CGG CTG AAG TCA TCC CAT GGA GAT GGC AGG CTC CTC Gln Val Ser Leu Arg Leu Lys Ser Ser His Gly Asp Gly Arg Leu Leu 625 630 635 640	2023
TGC GGG GCT ACG CTC CTG AGT AGC TGC TGG GTC CTC ACA GCA GCA CAC Cys Gly Ala Thr Leu Leu Ser Ser Cys Trp Val Leu Thr Ala Ala His 645 650 655	2071
TGT TTC AAG AGG TAT GGC AAC AGC ACT AGG AGC TAT GCT GTT AGG GTT Cys Phe Lys Arg Tyr Gly Asn Ser Thr Arg Ser Tyr Ala Val Arg Val 660 665 670	2119

GGA GAT TAT CAT ACT CTG GTA CCA GAG GAG TTT GAG GAA GAA ATT GGA Gly Asp Tyr His Thr Leu Val Pro Glu Glu Phe Glu Glu Glu Ile Gly 675 680 685	2167
GTT CAA CAG ATT GTG ATT CAT CGG GAG TAT CGA CCC GAC CGC AGT GAT Val Gln Gln Ile Val Ile His Arg Glu Tyr Arg Pro Asp Arg Ser Asp 690 695 700	2215
TAT GAC ATA GCC CTG GTT AGA TTA CAA GGA CCA GAA GAG CAA TGT GCC Tyr Asp Ile Ala Leu Val Arg Leu Gln Gly Pro Glu Glu Gln Cys Ala 705 710 715 720	2263
AGA TTC AGC AGC CAT GTT TTG CCA GCC TGT TTA CCA CTC TGG AGA GAG Arg Phe Ser Ser His Val Leu Pro Ala Cys Leu Pro Leu Trp Arg Glu 725 730 735	2311
AGG CCA CAG AAA ACA GCA TCC AAC TGT TAC ATA ACA GGA TGG GGT GAC Arg Pro Gln Lys Thr Ala Ser Asn Cys Tyr Ile Thr Gly Trp Gly Asp 740 745 750	2359
ACA GGA CGA GCC TAT TCA AGA ACA CTA CAA GCA GCC ATT CCC TTA Thr Gly Arg Ala Tyr Ser Arg Thr Leu Gln Ala Ala Ile Pro Leu 755 760 765	2407
CTT CCT AAA AGG TTT TGT GAA GAA CGT TAT AAG GGT CGG TTT ACA GGG Leu Pro Lys Arg Phe Cys Glu Glu Arg Tyr Lys Gly Arg Phe Thr Gly 770 775 780	2455
AGA ATG CTT TGT GCT GGA AAC CTC CAT GAA CAC AAA CGC GTG GAC AGC Arg Met Leu Cys Ala Gly Asn Leu His Glu His Lys Arg Val Asp Ser 785 790 795 800	2503
TGC CAG GGA GAC AGC GGA CCA CTC ATG TGT GAA CGG CCC GGA GAG Cys Gln Gly Asp Ser Gly Gly Pro Leu Met Cys Glu Arg Pro Gly Glu 805 810 815	2551
AGC TGG GTG GTG TAT GGG GTG ACC TCC TGG GGG TAT GGC TGT GGA GTC Ser Trp Val Val Tyr Gly Val Thr Ser Trp Gly Tyr Gly Cys Gly Val 820 825 830	2599
AAG GAT TCT CCT GGT GTT TAT ACC AAA GTC TCA GCC TTT GTA CCT TGG Lys Asp Ser Pro Gly Val Tyr Thr Lys Val Ser Ala Phe Val Pro Trp 835 840 845	2647
ATA AAA AGT GTC ACC AAA CTG TAA TTCTTCATGG AAACCTCAA GCAGCATT Ile Lys Ser Val Thr Lys Leu * 850 855	2700
AAACAAATGG AAAACTTGAA ACCCCCCACTA TTAGCACTCA GCAGAGATGA CAACAAATGG	2760
CAAGATCTGT TTTTGCTTTG TGTTGTGGTA AAAAATTGTG TACCCCCCTGC TGCTTTGAG	2820
AAATTTGTGA ACATTTTCAG AGGCCTCAGT GTAGTGGAAAG TGATAATCCT TAAATGAACA	2880
TTTTCTACCC TAATTCACT GGAGTGACTT ATTCTAACGCC TCATCTATCC CCTACCTATT	2940

TCTCAAAATC ATTCTATGCT GATTTACAA AAGATCATT TTACATTGA ACTGAGAAC 3000
CCTTTAATT GAATCAGTGG TGTCTGAAAT CATATTAAAT ACCCACATT GACATAAATG 3060
CGGTACCCTT TACTACACTC ATGAGTGGCA TATTTATGCT TAGGTCTTTT CAAAAGACTT 3120
GACAAGAAAT CTTCATATTC TCTGTAGCCT TTGTCAAGTG AGGAAATCAG TGGTTAAAGA 3180
ATTCCACTAT AAACTTTAG GCCTGAATAG GAGTAGTAAA GCCTCAAGGA CATCTGCCTG 3240
TCACAATATA TTCTCAAAGT GATCTGATAT TTGGAAACAA GTATCCTGT TGAGTACCAA 3300
GTGCTACAGA AACCATATAAGA TAAAAAATAC T TCTACCTAC AGCGTGCCCCG 3350

(1) INFORMATION ABOUT THE COMPOUND OF THE FORMULA II (Neurotrypsin of the mouse)

5 (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2376 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single strand
- (D) TOPOLOGY: linear

10 (ii) MOLECULE TYPE: cDNA to mRNA

(vi) ORIGINAL SOURCE:

15 (A) ORGANISM: Mus musculus
(D) DEVELOPMENT STAGE: postnatal day 10
(F) TISSUE TYPE: brain
(G) CELL TYPE: neurons

20 (vii) IMMEDIATE SOURCE:

(A) LIBRARY: mouse brain cDNA library in the lambda Uni-ZAP-XR vector, oligo (dT)-primed, from Balb c mice, postnatal day 20,
Cat. No.. 937 319; Stratagene, La Jolla, CA, USA

25

(B) CLONE: cDNA clone no. 16

(vii) IMMEDIATE SOURCE:

30

(A) LIBRARY: mouse brain cDNA library in the Lambda gt10 vector,
oligo(dT)- and random-primed, embryonic day 15,
Cat. No. ML 3002a; Clontech, Palo Alto, CA, USA

35 (B) CLONE: cDNA clone #25

(ix) FEATURE:

(A) NAME/KEY: signal peptide

5 (B) LOCATION: 24 .. 86

(ix) FEATURE:

10 (A) NAME/KEY: mature peptide

(B) LOCATION: 87 .. 2306

(ix) FEATURE:

15

(A) NAME/KEY: coding sequence

(B) LOCATION: 24 .. 2306

20 (ix) FEATURE:

(A) NAME/KEY: proline-rich, basic segment

(B) LOCATION: 90 .. 275

25

(ix) FEATURE:

(A) NAME/KEY: Kringle domain

(B) LOCATION: 276 .. 494

30

(ix) FEATURE:

(A) NAME/KEY: SRCR domain 1

35 (B) LOCATION: 519 .. 824

(ix) FEATURE:

5 (A) NAME/KEY: SRCR domain 2
(B) LOCATION: 840 .. 1142

10 (ix) FEATURE:

(A) NAME/KEY: SRCR domain 3
(B) LOCATION: 1179 .. 1484

15 (ix) FEATURE:

(A) NAME/KEY: proteolytic domain
(B) LOCATION: 1536 .. 2306

20 (ix) FEATURE:

(A) NAME/KEY: histidine of the catalytic triade
(B) LOCATION: 1707 .. 1709

25

(ix) FEATURE:

30 (A) NAME/KEY: aspartic acid of the catalytic triade
(B) LOCATION: 1857 .. 1859

(ix) FEATURE:

35 (A) NAME/KEY: serine of the catalytic triade

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(B) LOCATION: 2154 .. 2156

(ix) FEATURE:

5 (A) NAME/KEY: polyA signal

(B) LOCATION: 2324 .. 2329 and 2331 .. 2336

(ix) FEATURE:

10 (A) NAME/KEY: polyA segment

(B) LOCATION: 2357 .. 2376

(ix) FEATURE:

15

(A) NAME/KEY: 3'UTR

(B) LOCATION: 2307 .. 2341 or 2307 .. 2356

20

(ix) FEATURE:

(A) NAME/KEY: 5'UTR

(B) LOCATION: 1 .. 23

Compound of the formula II (neurotrypsin of the mouse)

GGACCACACT CGGCGCCGCA GCC ATG GCG CTC GCC CGC TGC GTG CTG GCT GTG	53
Met Ala Leu Ala Arg Cys Val Leu Ala Val	
-20	-15
ATT TTA GGG GCA CTG TCT GTA GTG GCC CGC GCT GAT CCG GTC TCG CGC	101
Ile Leu Gly Ala Leu Ser Val Val Ala Arg Ala Asp Pro Val Ser Arg	
-10	-5
TCT CCC CTT CAC CGC CCG CAT CCG TCC CCA CCG CGT TCC CAA CAC GCG	149
Ser Pro Leu His Arg Pro His Pro Ser Pro Pro Arg Ser Gln His Ala	
10	15
CAC TAC CTT CCC AGC TCG CGG CGG CCA CCC AGG ACC CCG CGC TTC CCG	197
His Tyr Leu Pro Ser Ser Arg Arg Pro Pro Arg Thr Pro Arg Phe Pro	
25	30
CTC CCG CTG CGG ATC CCC GCT GCC CAG CGC CCG CAG GTC CTC AGC ACC	245
Leu Pro Leu Arg Ile Pro Ala Ala Gln Arg Pro Gln Val Leu Ser Thr	
40	45
GGG CAC ACG CCC CCG ACG ATT CCA CGC CGC TGC GGG GCA GGA GAG TCG	293
Gly His Thr Pro Pro Thr Ile Pro Arg Arg Cys Gly Ala Gly Glu Ser	
55	60
TGG GGC AAT GCC ACC AAC CTC GGC GTC CCG TGT CTA CAC TGG GAC GAG	341
Trp Gly Asn Ala Thr Asn Leu Gly Val Pro Cys Leu His Trp Asp Glu	
70	75
GTG CCG CCC TTC CTG GAG CGG TCG CCC CCG GCC AGT TGG GCT GAG CTG	389
Val Pro Pro Phe Leu Glu Arg Ser Pro Pro Ala Ser Trp Ala Glu Leu	
90	95
CGA GGG CAG CCG CAC AAC TTC TGC CGG AGC CCG GAT GGC TCG GGC AGA	437
Arg Gly Gln Pro His Asn Phe Cys Arg Ser Pro Asp Gly Ser Gly Arg	
105	110
CCT TGG TGC TTC TAT CCG AAT GCC CAG GGC AAA GTA GAC TGG GGC TAC	485
Pro Trp Cys Phe Tyr Arg Asn Ala Gln Gly Lys Val Asp Trp Gly Tyr	
120	125
TGC GAT TGT GGT CAA GGC CCG GCG TTG CCC GTC ATT CGC CTT GTT GGT	533
Cys Asp Cys Gly Gln Gly Pro Ala Leu Pro Val Ile Arg Leu Val Gly	
135	140
GGG AAC AGT GGG CAT GAA GGT CGA GTG GAG CTG TAC CAC GCT GGC CAG	581
Gly Asn Ser Gly His Glu Gly Arg Val Glu Leu Tyr His Ala Gly Gln	
150	155
TGG GGG ACC ATC TGT GAC GAC CAA TGG GAC AAT GCA GAC GCA GAC GTC	629
Trp Gly Thr Ile Cys Asp Asp Gln Trp Asp Asn Ala Asp Ala Asp Val	
170	175
ATC TGT AGG CAG CTG GGG CTC AGT GGC ATT GCC AAA GCA TGG CAT CAG	677
Ile Cys Arg Gln Leu Gly Leu Ser Gly Ile Ala Lys Ala Trp His Gln	
185	190
195	

GCA CAT TTT GGG GAA GGA TCT GGC CCA ATA TTG TTG GAT GAA GTA CGC Ala His Phe Gly Glu Gly Ser Gly Pro Ile Leu Leu Asp Glu Val Arg 200 205 210	725
TGC ACC GGA AAC GAG CTG TCA ATT GAG CAA TGT CCA AAG AGT TCC TGG Cys Thr Gly Asn Glu Leu Ser Ile Glu Gln Cys Pro Lys Ser Ser Trp 215 220 225	773
GGC GAA CAT AAC TGT GGC CAT AAA GAA GAT GCT GGA GTG TCT TGT GTT Gly Glu His Asn Cys Gly His Lys Glu Asp Ala Gly Val Ser Cys Val 230 235 240 245	821
CCT CTA ACA GAT GGT GTC ATC AGA CTG GCA GGA GGA AAA AGT ACC CAT Pro Leu Thr Asp Gly Val Ile Arg Leu Ala Gly Gly Lys Ser Thr His 250 255 260	869
GAA GGT CGC CTG GAG GTC TAC TAC AAG GGG CAG TGG GGG ACA GTC TGT Glu Gly Arg Leu Glu Val Tyr Tyr Lys Gly Gln Trp Gly Thr Val Cys 265 270 275	917
GAT GAT GCC TGG ACT GAG ATG AAC ACA TAC GTG GCT TGT CGA CTG CTG Asp Asp Gly Trp Thr Glu Met Asn Thr Tyr Val Ala Cys Arg Leu Leu 280 285 290	965
GGA TTT AAA TAC GGC AAA CAG TCC TCT GTG AAC CAT TTT GAT GGC AGC Gly Phe Lys Tyr Gly Lys Gln Ser Ser Val Asn His Phe Asp Gly Ser 295 300 305	1013
AAC AGG CCC ATA TGG CTG GAT GAC GTC AGC TGC TCA GGA AAA GAA GTC Asn Arg Pro Ile Trp Leu Asp Asp Val Ser Cys Ser Gly Lys Glu Val 310 315 320 325	1061
AGC TTC ATT CAG TGT TCC AGG AGA CAG TGG GGA AGG CAT GAC TGC AGC Ser Phe Ile Gln Cys Ser Arg Arg Gln Trp Gly Arg His Asp Cys Ser 330 335 340	1109
CAT AGA GAA GAT GTG GGC CTC ACC TGC TAT CCT GAC AGC GAT GGA CAT His Arg Glu Asp Val Gly Leu Thr Cys Tyr Pro Asp Ser Asp Gly His 345 350 355	1157
AGG CTT TCT CCA GGT TTT CCC ATC AGA CTA GTG GAT GGA GAG AAT AAG Arg Leu Ser Pro Gly Phe Pro Ile Arg Leu Val Asp Gly Glu Asn Lys 360 365 370	1205
AAG GAA GGA CGA GTG GAG GTT TTT GTC AAT GGC CAA TGG GGA ACA ATC Lys Glu Gly Arg Val Glu Val Phe Val Asn Gly Gln Trp Gly Thr Ile 375 380 385	1253
TGC GAT GAC GGA TGG ACC GAT AAG CAT GCA GCT GTG ATC TGC CGG CAA Cys Asp Asp Gly Trp Thr Asp Lys His Ala Ala Val Ile Cys Arg Gln 390 395 400 405	1301
CTT GGC TAT AAG GGT CCT GCC AGA GCA AGG ACT ATG GCT TAT TTT GGG Leu Gly Tyr Lys Gly Pro Ala Arg Ala Arg Thr Met Ala Tyr Phe Gly 410 415 420	1349
GAA GGA AAA GGC CCC ATC CAC ATG GAT AAT GTG AAG TGC ACA GGA AAT Glu Gly Lys Gly Pro Ile His Met Asp Asn Val Lys Cys Thr Gly Asn 425 430 435	1397

GAG AAG GCC CTG GCT GAC TGT GTC AAA CAA GAC ATT GGA AGG CAC AAC Glu Lys Ala Leu Ala Asp Cys Val Lys Gln Asp Ile Gly Arg His Asn 440 445 450	1445
TGC CGC CAC AGT GAG GAT GCA GGA GTC ATC TGT GAC TAT TTA GAG AAG Cys Arg His Ser Glu Asp Ala Gly Val Ile Cys Asp Tyr Leu Glu Lys 455 460 465	1493
AAA GCA TCA AGT AGT GGT AAT AAA GAG ATG CTC TCA TCT GGA TGT GGA Lys Ala Ser Ser Ser Gly Asn Lys Glu Met Leu Ser Ser Gly Cys Gly 470 475 480 485	1541
CTG AGG TTA CTG CAC CGT CGG CAG AAA CGG ATC ATT GGT GGG AAC AAT Leu Arg Leu Leu His Arg Arg Gln Lys Arg Ile Ile Gly Gly Asn Asn 490 495 500	1589
TCT TTA AGG GGT GCC TGG CCT TGG CAG GCT TCC CTC AGG CTG AGG TCG Ser Leu Arg Gly Ala Trp Pro Trp Gln Ala Ser Leu Arg Leu Arg Ser 505 510 515	1637
GCC CAT GGA GAC GGC AGG CTG CTT TGT GGA GCT ACC CTT CTG AGT AGC Ala His Gly Asp Gly Arg Leu Leu Cys Gly Ala Thr Leu Leu Ser Ser 520 525 530	1685
TGC TGG GTC CTG ACA GCT GCA CAC TGC TTC AAA AGG TAC GGA AAC AAC Cys Trp Val Leu Thr Ala Ala His Cys Phe Lys Arg Tyr Gly Asn Asn 535 540 545	1733
TCG AGG AGC TAT GCA GTT CGA GTT GGG GAT TAT CAT ACT CTG GTC CCA Ser Arg Ser Tyr Ala Val Arg Val Gly Asp Tyr His Thr Leu Val Pro 550 555 560 565	1781
GAG GAG TTT GAA CAA GAA ATA GGG GTT CAA CAG ATT GTG ATT CAC AGG Glu Glu Phe Glu Gln Glu Ile Gly Val Gln Gln Ile Val Ile His Arg 570 575 580	1829
AAC TAC AGG CCA GAC AGA AGC GAC TAT GAC ATT GCC CTG GTT AGA TTG Asn Tyr Arg Pro Asp Arg Ser Asp Tyr Asp Ile Ala Leu Val Arg Leu 585 590 595	1877
CAA GGA CCA GGG GAG CAA TGT GCC AGA CTA AGC ACC CAC GTT TTG CCA Gln Gly Pro Gly Glu Gln Cys Ala Arg Leu Ser Thr His Val Leu Pro 600 605 610	1925
GCC TGT TTA CCT CTA TGG AGA GAG AGG CCA CAG AAA ACA GCC TCC AAC Ala Cys Leu Pro Leu Trp Arg Glu Arg Pro Gln Lys Thr Ala Ser Asn 615 620 625	1973
TGT CAC ATA ACA GGA TGG GGA GAC ACA GGT CGT GCC TAC TCA AGA ACT Cys His Ile Thr Gly Trp Gly Asp Thr Gly Arg Ala Tyr Ser Arg Thr 630 635 640 645	2021
CTA CAA CAA GCT GCT GTG CCT CTG TTA CCC AAG AGG TTT TGT AAA GAG Leu Gln Gln Ala Ala Val Pro Leu Leu Pro Lys Arg Phe Cys Lys Glu 650 655 660	2069
AGG TAC AAG GGA CTA TTT ACT GGG AGA ATG CTC TGT GCT GGG AAC CTC Arg Tyr Lys Gly Leu Phe Thr Gly Arg Met Leu Cys Ala Gly Asn Leu 665 670 675	2117

CAA GAA GAC AAC CGT GTG GAC AGC TGC CAG GGA GAC AGT GGA GGA CCA	2165
Gln Glu Asp Asn Arg Val Asp Ser Cys Gln Gly Asp Ser Gly Gly Pro	
680 685 690	
CTC ATG TGT GAA AAG CCT GAT GAG TCC TGG GTT GTG TAT GGG GTG ACT	2213
Leu Met Cys Glu Lys Pro Asp Glu Ser Trp Val Val Tyr Gly Val Thr	
695 700 705	
TCC TGG GGG TAT GGA TGT GGA GTC AAA GAC ACT CCT GGA GTT TAT ACC	2261
Ser Trp Gly Tyr Gly Cys Gly Val Lys Asp Thr Pro Gly Val Tyr Thr	
710 715 720 725	
AGA GTC CCC GCT TTT GTA CCT TGG ATA AAA AGT GTC ACC AGT CTG	2306
Arg Val Pro Ala Phe Val Pro Trp Ile Lys Ser Val Thr Ser Leu	
730 735 740	
TAACCTTATGG AAAGCTCAAG AAATAGTAAA ACAGTAACTA TTCAGTCTTC AAAAAAAA	2366
AAAAAAAAAA	2376